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PSEUDOMONAS AERUGINOSA: INFECTION AND IMMUNITY

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Pseudomonas aeruginosa is a major cause of infections in Western society, being the causative agent of about 200 000 nosocomial infections per year in North America. Such infections tend to be rapidly progressing and pose serious risks to patients. However, P. aeruginosa can also cause long-term (up to 10 years or more) chronic infections in the lungs of patients with cystic fibrosis, and the onset of P. aeruginosa infection signals the eventual deterioration of lung function in such patients, leading to death from respiratory failure. P. aeruginosa shows high intrinsic antibiotic resistance. While there are antibiotics that show initial success against Pseudomonas infections, even modest mutations lead to clinically significant resistance, making development of antibiotic resistance common and successful treatment difficult. For this reason considerable effort has been expended in seeking vaccine or immunotherapeutic solutions to such infections. At the same time, diagnosis and epidemiology, each of which use specific antigens, has become important in monitoring the onset and course of infection and can influence the eventual success of therapy.

Process of infection

P. aeruginosa causes rapidly progressing infections in animals and humans. Generally speaking, these infections only occur in those animals and people which have a reduced capacity to defend against infection (or at body sites that have intrinsically weaker defenses). Thus, in animal models, mild alcohol burns or neutropenia induced by cyclophosphamide lead to a reduction in the LD₅₀ (the concentration of bacteria that kill 50% of animals) by up to 100 000-fold. Similarly, the types of human conditions which tend to predispose towards *P. aeruginosa* infections include neutropenia induced by anticancer drugs, burns, major surgery, steroid therapy, etc. In such patients the likelihood of *P. aeruginosa* infection increases with length of hospital stay.

P. aeruginosa produces a multitude of virulence

| Anugen | Characteristics/function | Specific antibodies/use | Immunogenicity |
|--|---|--|--|
| Cell associated: | | | |
| Outer membrane protein OprF | Major pore-forming protein; structural role | Opsonic and protective; mAbs used in structural studies, diagnostics; antigenically conserved | Strong Ab response |
| OprL (= H2) | Peptidoglycan-associated lipoprotein (PAL); structural role | mAb reveals conserved antigen in type I Pseudomonadaceae | Strong Ab response |
| Oprl | Braun lipoprotein | Protective; mAbs reveal antigenic conservation | Strong Ab response |
| Iron-regulated outer membrane proteins | Transport of iron-siderophore complexes | | Ab response accompanies chronic infection |
| Variant lipopolysaccharide (LPS) A-band (D-rhamnan) | Common antigen, induces influx of PMN | Not neutralizing or opsonic | Ab response appears late ir patients with chronic |
| LPS B-band (O antigen of LPS) | O-polysaccharide of conventional LPS; forms the basis for serotype-specificity | Serotype-specifically opsonic and protective; mAb/polyclonals against each serotype produced; useful in serotyping and epidemiology | Strong Ab response |
| PS core polysaccharide | Conserved region which links O antigen polymers to lipid A | Cross-reactive; inner core epitope exposed on cell surface | Strong Ab response |
| .PS lipid A | Anchors LPS in the outer membrane; endotoxic moiety.– causes fever and septic shock | Ab broadly cross-reactive even with other species; reacts poorly with intact cells | Weak immunogen, includes <i>P. aeruginosa-</i> specific response |
| /lucoid exopolysaccharide/alginate | Exopolysaccharide in the form of loose slime; associated with biofilm/microcolony formation | Both conserved and selective Abs available | Weak unless conjugated to protein |
| lagella | Motility; chemotaxis and invasion | Two major serogroups (type a with 4 subgroups and type b); opsonic; inhibit motility; protective; mAbs available | Strong Ab response |
| | Adherence and initial colonization | About 14 types; antigenically variable; Neutralizing and passively protective; many mAbs available | Weak to intermediate Ab response |
| bosomes | Protein synthesis | Protective property not confirmed; may be due to contamination | |
| tracellular proteins: | | contamination | |
| astase | Degrades connective tissues. Tissue invasion | | Observed in chronic |
| otoxin A | Inhibition of host cell protein synthesis | Cross-reactive neutralizing activity | Strong Ab response |
| oenzyme S | Inhibition of host cell protein synthesis | Neutralizing activity | |

Table 1 Immunogenicity and antibody response to major antigens of Pseudomonas aeruginosa

factors including the extracellular toxins exotoxin A and exoenzyme S, a periplasmic cytotoxin, two secreted proteases elastase and alkaline protease, secreted lipase, phospholipase and heat-stable hemo-

lysin (also called rhamnolipid), a neurominidase, slime glycolipoprotein, mucoid exopolysaccharide, and endotoxin. Each of these has been shown in animal models to have a role in pathogenesis, making *Pseudomonas* pathogenesis truly multideterminant, and in many cases in those same animal models, immune intervention directed against the virulence factor in question will reduce the impact of, or even prevent infection. In rapidly progressing infections, which can be common in nosocomial situations, perhaps the most important characteristic of *P. aeruginosa* is its ability to grow rapidly *in vivo*, with a doubling time equivalent to that observed on a nutrient medium *in vitro*. Thus in a relatively short time (24–36 h) such infections can become overwhelming.

In the more complex situation posed by the chronic infections observed with cystic fibrosis patients, it is still not entirely clear why this disease predisposes a patient towards a long-term, localized lung infection by a bacterium which normally causes rapidly developing invasive infections. The suggestion is that localized defects lead to establishment of infection. One newer concept is that the high salt concentration induced by the mutation of the cystic fibrosis transmembrane regulator (CFTR) protein, renders ineffective the natural antimicrobial defense peptides in the lung.

antibodies (mAbs) have been raised against these (and other) antigens and have been useful in demonstrating the varied levels of serological conservation in these antigens. Thus, there are 20 lipopolysaccharide (LPS) O-serotypes, about 14 pilin antigenic variants and about five flagellin variants (representative of two major serotypes) in P. aeruginosa. This makes these antigens ideal as targets for epidemiological studies. On the other hand, the outer membrane proteins OprF, OprL and OprI, LPS rough core and lipid A, exoantigens and mucosal exoplysaccharide have only one or two variant antigenic types. Such antigens therefore have substantial potential as targets for diagnosing P. aeruginosa infections and most of these are or have been considered as vaccines. The mAbs raised against *P. aeruginosa* antigens have also proven to be excellent tools for studying these antigens and have assisted in the study of the structure, function, molecular genetics and immunology of these proteins.

Immunogenicity and immunotherapy

Antigenicity

As indicated in Table 1, there are a wide range of antigens that have been studied in detail in *P. aeruginosa*. Specific antisera and in many cases monoclonal

All of the antigens listed in **Table 1** have been shown to elicit antibodies in animal models, cystic fibrosis patients with *P. aeruginosa*-infected lungs, and/or humans with burn wound infections. This has provided the rationale for pursuit of vaccine and passive immunotherapy studies (**Table 2**). In many cases,

Table 2 Recent immunotherapeutic or immunoprophylactic preparations against Pseudomonas aeruginosa

| Vaccine | Efficacy assessment and other remarks | |
|---|---|--|
| Octavalent LPS O-polysaccharide-toxin A conjugate vaccine | Engendered IgG response that was protective. However human intravenous immunoglobulins versus this vaccine did not show statistically significant protection in human trials | |
| Alginate-toxin A conjugate vaccine | Serum antibodies from rabbits were opsonic and promoted killing of <i>Pseudomonas aeruginosa</i> , and had neutralizing activities towards toxin A | |
| Anti-idiotype antibodies | Induced LPS-specific response in mice | |
| OprF-OprI hybrid vaccine | Significant antibody titers in human volunteers after immunization; protective in immunosuppressed mouse model study | |
| Alginate vaccine | Different sizes of alginate in preparation had variable effect in eliciting opsonic antibody response; large molecular size antigen preparation was tolerated well in volunteers; human trials apparently abandoned | |
| Synthetic peptides of OprF epitopes | Linear B cell epitopes of OprF-conjugated with carrier protein as vaccine. Elicited protective response | |
| ScFv of OprF-specific antibody | Modern technology; single-chain antibody specific against an OprF domain. Applicable for epitope mapping and further antibody engineering | |
| Killed whole bacteria delivered mucosally | Demonstrated heterologous protection in mice | |
| Pilin peptides | Limited cross-reactivity observed but attempts are being made to improve this by peptide engineering; protective in mice | |
| Human mAbs | Five serotype-specific anti-LPS O-antigen human mAbs were opsonophagocytic and protective in mice; three flagella specific human mAbs were protective in mice | |
| Human-Murine heterohybridomas | Human antibody specific against <i>Pseudomonas aeruginosa</i> , scale-up production successful, but efficacy is not defined at present | |

injection of the antigen into animals leads to a sufficient production of antibodies that promote opsonophagocytosis and protection against subsequent infections. Since P. aeruginosa is generally considered to be an extracellular infection, there is a general consensus that an antibody response is required for protection. However, this is not necessarily sufficient for protection since the lungs of patients with cystic fibrosis continue to remain chronically colonized despite exuberant serum antibody responses to several antigens. Furthermore, there are few circumstances in which P. aeruginosa infections are truly predictable, making active vaccination a less attractive prospect. For this reason passive protections with hyperimmune intravenous immunoglobulins from volunteers immunized with a Pseudomonas antigen has been pursued. Unfortunately, the most recent of these studies using an intravenous immunoglobulin with strong titers to the most predominant LPS serotypes and to exotoxin A, was abandoned due to its inability to show statistically significant protection in humans at the dosage used combined with a less favorable adverse reaction profile. While it is too early to discard the possibility for success with one of the other vaccine strategies being pursued, it may require a novel approach to yield success.

See also: Bacteria, immunity to; Bacterial cell walls.

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